A simple model for the eukaryotic cell cycle

Andrea Ciliberto



The cell division cycle





Kohn, Mol. Biol. Cell., 1999

How did we get to this mess??

Dominoes and Clocks: The Union of Two Views of the Cell Cycle

ANDREW W. MURRAY AND MARC W. KIRSCHNER

We review the recent advances in understanding transitions within the cell cycle. These have come from both genetic and biochemical approaches. We discuss the phylogenetic conservation of the mechanisms that induce mitosis and their implications for other transitions in the cell cycle.

THE CELL CYCLE IS THE SET OF EVENTS THAT IS RESPONSIBLE for the duplication of the cell. The recent advances in our understanding of the cell cycle have come from two approaches. Geneticists attempted to understand the cell cycle by analyzing mutations that arrested the cell cycle of somatic cells at specific points, whereas embryologists and physiologists examined natural points of cell cycle arrest and the agents that induced the embryonic cell cycle to proceed.

The genetic approach to the somatic cell cycle evolved from prokaryotic genetics in the 1950s and 1960s. With genetics, researchers successfully explained complicated processes, such as phage morphogenesis, as a linear sequence of events. The most extreme models of these processes suggested that they would resemble metabolic pathways: the initiation of each step in the pathway would be dependent on the completion of the preceding step, because the product of the earlier step was the substrate for the latter one; specific genes were assumed to execute each step. When this approach was applied to yeast, first in the budding yeast by Hartwell and his colleagues (1) and later in the fission yeast by Nurse and his colleagues (2), the result was a description of the cell cycle as a set of dependent reactions. The basis of this dependency is discussed in the accompanying review by Hartwell and Weinert (3). The physiological and embryological approach was championed by researchers who favored marine and amphibian eggs. They argued that eggs and oocytes were the simplest systems for studying the basic processes of the cell cycle, because they were specialized for rapid cell division. The result of their investigations was a description of the cell cycle as a biochemical machine that oscillated between two states, mitosis and interphase, and whose oscillations were independent of the completion of many of the cell cycle events. Initially the two views of the cell cycle, one as a set of dependent reactions (the domino theory) and the other as a biochemical oscillator (the clock theory), seemed incompatible.

The cell fusion experiments of Rao and Johnson (4) supported both points of view. The fusion of cells in mitosis with cells in any other state induced some form of mitotic response in the interphase nucleus and supported the embryological model of distinct mitotic and interphase cytoplasmic states, with the mitotic state dominant over all interphase states (4). Fusion experiments, however, also supported the idea of a dependent cell cycle, since in any fusion between two interphase cells at different stages of the cell cycle, the advanced nucleus waits for the completion of events in the retarded nucleus before progressing in the cell cycle (4).

In this review we discuss recent evidence from both traditions that has led to a unified view of the eukaryotic cell cycle. This synthesis suggests that a single biochemical mechanism underlies the cell cycle in all cukaryotic organisms. We have concentrated on the reactions that regulate progress through the cell cycle and do not discuss the mechanism of individual cell cycle events such as DNA synthesis or nuclear envelope breakdown and reformation.

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Xenopus and the clock paradigm



Cell mass decreases during early divisions

Alberts et a., Molecular Biology of the Cell.2002

In Xenopus oscillations progress independently of DNA presence and cell cycle events

Autonomous oscillations!



Alberts et a., Molecular Biology of the Cell.2002

MPF, the mitosis promoting factor





Murray and Kirschner, Science, 1989

MPF is a heterodimer



cyclin dependent kinase cyclin (regulatory subunit)

Only cyclin synthesis and degradation are required for *Xenopus* early cycles.



Alberts et a., Molecular Biology of the Cell.2002

CDK is activated by cyclin binding and once activated it induces cyclin degradation



But something else must be at work...



Cyclin threshold



Solomon et al, Cell, 1990

Yeast and the domino paradigm



Balanced growth and division



Cell division cycle (cdc) mutants are temperature sensitive



Figure 5. Normal cells and cdc mutant cells several hours after incubation at the restrictive temperature. (A) wild type, (B) cdc8 (C) cdc24 (D) cdc10.



Figure 6. A pathway of gene controlled events in the *S. cerevisiae* cell cycle. Numbers refer to *cdc* genes. Abbreviations are: iDS, initiation of DNA synthesis, DS, DNA synthesis, mND, medial nuclear division; IND, late nuclear division; BE, bud emergence; NM, nuclear migration; CK, cytokinesis; CS cell separation; MF mating factor. Reprinted from ref 7 with permission.

Hartwell, Genetics, 1991



Alberts et a., Molecular Biology of the Cell.2002

Wee1 controls a rate limiting step in the cell cycle

Cell division and cell growth are coupled



Nurse, Noble lecture, 2000

Basic cell cycle properties

- Cell physiology-

- Coupling of mass growth and cell division.
- Once the cell enters the cycle, it is committed to finish it: irreversibility.
- The cell halts during cell cycle progression if something has gone wrongly.

-Molecular network-

- -Oscillations of MPF drive cells into and out of mitosis.
- Cdc28 activity is controlled by Wee1 (negative) and Cdc25 (positive).

Dominoes and clocks: Cdc28 is the budding yeast homologous of MPF's catalytic subunit



Figure 6. A pathway of gene controlled events in the *S. cerevisiae* cell cycle. Numbers refer to *cdc* genes. Abbreviations are: iDS, initiation of DNA synthesis, DS, DNA synthesis, mND, medial nuclear division; IND, late nuclear division; BE, bud emergence; NM, nuclear migration; CK, cytokinesis; CS cell separation; MF mating factor. Reprinted from ref 7 with permission.



Phosphorylation as well as cyclin binding controls MPF activity



Phosphorylation as well as cyclin binding controls MPF activity



Isolation and analysis of a positive feedback: the network...







Notice, here no cyclin synthesis, no cyclin degradation!!

...and the physiology



Solomon et al, Cell, 1990

Part II Standard laws of biochemical kinetics applied to molecular networks

Law of Mass Action: forward reaction

pMPF → MPF

$$\frac{dMPF}{dt} = k_{a} \cdot pMPF$$

$$pMPF = MPF_{tot} - MPF$$

$$\frac{dMPF}{dt} = k_{a} \cdot (MPF_{tot} - MPF)$$

$$Steady State solution (MPF^{SS})$$

$$\frac{dMPF}{dt} = 0$$

$$MPF^{SS} = MPF_{tot}$$

Notice: no dimer, only MPF. Cdk is supposed to be present in excess throughout the cycle. Increasing MPF total mimics an increase in cyclin total.





Law of Mass Action: reversible reaction



Steady State solution $\frac{dMPF}{dt} = 0$ $MPF^{SS} = \frac{k_a \cdot MPF_{tot}}{k_a + k_i}$







Law of Mass Action: catalyzed reversible reaction





Nullclines



What happens if MPF total increases?



Michaelis-Menten: forward reaction

Wee1P → Wee1

dWee1	kwa · Wee1P
dt	$\overline{J + Wee1P}$
dWee1	$\underline{k_{wa}} \cdot (Wee1_{tot} - Wee1)$
dt	$\overline{J + (Wee1_{tot} - Wee1)}$

Steady State solution $\frac{dWee1}{dt} = 0$ $Wee1^{SS} = Wee1_{tot}$



Michaelis-Menten: reversible reaction



Proc. Natl. Acad. Sci. USA Vol. 78, No. 11, pp. 6840-6844, November 1981 Biochemistry

An amplified sensitivity arising from covalent modification in biological systems

(protein modification/metabolic regulation/switch mechanism/enzyme cascades)

ALBERT GOLDBETER[†] AND DANIEL E. KOSHLAND, JR. Department of Biochemistry, University of California, Berkeley, California 94720 Contributed by Daniel E. Koshland, Jr., August 11, 1981

if [enzym1_{TOT}], [enzyme2_{TOT}] << [Wee1_{TOT}]

$$k_{wa}$$
=[enzyme1_{TOT}] k_2
 k_{wi} =[enzyme2_{TOT}] k_4

dWee1_	$\frac{k_{\text{wa}} \cdot (\text{Wee1}_{\text{tot}} - \text{Wee1})}{k_{\text{tot}} - \text{Wee1}}$	$k_{wi} \cdot Wee1$
dt	$J + Wee1_{tot} - Wee1$	J + Wee1
	production	elimination

Michaelis-Menten: reversible reaction



Nullclines



Phase plane analysis



$$\frac{dMPF}{dt} = k_a \cdot (MPF_{tot} - MPF) - k_i \cdot MPF \cdot Wee1$$

$$\frac{dWee1}{dt} = \frac{k_{wa} \cdot (Wee1_{tot} - Wee1)}{J + Wee1_{tot} - Wee1} - \frac{k_{wi} \cdot Wee1}{J + Wee1}$$





How does MPF increases with Cyclin total?

$$MPF^{SS} = \frac{k_{a} \cdot MPF_{tot}}{k_{i} \cdot Wee1 + k_{a}}$$



MPFtot

Not quite the same!

	1	2	3	4	5	6	7	8	9	10
nM CYCLIN:				1500	460	150	46	15	4.6	1.5
				-			1		r ist	- 31.
H1 KINASE:	104	17	16	325	249	139	188	20	16	14
MPF:	440	<30	<30	880	680	520	360	<30	<30	<30

Solomon et al, Cell, 1990



Michaelis-Menten: catalyzed reversible reaction



$\underline{k_{wa}} \cdot (Wee1_{tot} - Wee1)$	$\underline{k_{wi}} \cdot Wee1 \cdot MPF$
$J + Wee1_{tot} - Wee1$	J+Wee1
production	elimination
	$=\frac{k_{wa} \cdot (Wee1_{tot} - Wee1)}{J + Wee1_{tot} - Wee1}$ production +

Nullclines



Phase plane analysis



Phase plane analysis





First solution, MPF wins, Wee1 loses





Third solution, both can win: hysteresis



How does MPF increases with Cyclin total?



Hysteresis in the Xenopus early cycles: simulation of an experimental result







What happens if cyclin total increases with cell mass?





NETWORK DYNAMICS AND CELL PHYSIOLOGY

John J. Tyson*, Kathy Chen* and Bela Novak‡

NATURE REVIEWS | MOLECULAR CELL BIOLOGY DECEMBER 2001 | VOLUME 2







Conclusion

-Same wiring in different organisms, combination of positive and negative feedbacks.

- In Xenopus early development, with large mass, the cell cycle is a limit cycle oscillator, the negative feedback plays the key role.

- Artificially, an additional mechanism of control emerges, based on a positive feedback loop.

- Both positive and negative feedbacks are at work in yeast. In these organisms, mass growth drives the cell cycle.

- Positive feedbacks introduce checkpoints and irreversibility in the cycle.

The negative feedback the capability to start a new process.

Can a biologist fix a radio?—Or, what I learned while studying apoptosis

CANCER CELL : SEPTEMBER 2002

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Figure 3. The tools used by biologists and engineers to describe processes of interest

A: The biologist's view of a radio. See Figure 2 and text for description of the indicated components. **B**: The engineer's view of a radio. (Please note that the circuit diagram presented is not that of the radio used in the study. The diagram of the radio was lost, which, in part, explains why the radio remains broken.)